

# The Identification of a Small Molecule Compound That Reduces HIV-1 Nef-Mediated Viral Infectivity Enhancement

Nopporn Chutiwitoonchai<sup>1</sup>, Masateru Hiyoshi<sup>1</sup>, Philip Mwimanzi<sup>1</sup>, Takamasa Ueno<sup>1</sup>, Akio Adachi<sup>2</sup>, Hirotaka Ode<sup>3</sup>, Hironori Sato<sup>3</sup>, Oliver T. Fackler<sup>4</sup>, Seiji Okada<sup>1</sup>, Shinya Suzu<sup>1\*</sup>

**1** Center for AIDS Research, Kumamoto University, Kumamoto, Japan, **2** Department of Microbiology, Institute of Health Biosciences, The University of Tokushima Graduate School, Tokushima, Japan, **3** Pathogen Genomics Center, National Institute of Infectious Diseases, Tokyo, Japan, **4** Department of Infectious Diseases, Virology, University of Heidelberg, Heidelberg, Germany

## Abstract

Nef is a multifunctional HIV-1 protein that accelerates progression to AIDS, and enhances the infectivity of progeny viruses through a mechanism that is not yet understood. Here, we show that the small molecule compound 2c reduces Nef-mediated viral infectivity enhancement. When added to viral producer cells, 2c did not affect the efficiency of viral production itself. However, the infectivity of the viruses produced in the presence of 2c was significantly lower than that of control viruses. Importantly, an inhibitory effect was observed with Nef<sup>+</sup> wild-type viruses, but not with viruses produced in the absence of Nef or in the presence of proline-rich PxxP motif-disrupted Nef, both of which displayed significantly reduced intrinsic infectivity. Meanwhile, the overexpression of the SH3 domain of the tyrosine kinase Hck, which binds to a PxxP motif in Nef, also reduced viral infectivity. Importantly, 2c inhibited Hck SH3-Nef binding, which was more marked when Nef was pre-incubated with 2c prior to its incubation with Hck, indicating that both Hck SH3 and 2c directly bind to Nef and that their binding sites overlap. These results imply that both 2c and the Hck SH3 domain inhibit the interaction of Nef with an unidentified host protein and thereby reduce Nef-mediated infectivity enhancement. The first inhibitory compound 2c is therefore a valuable chemical probe for revealing the underlying molecular mechanism by which Nef enhances the infectivity of HIV-1.

**Citation:** Chutiwitoonchai N, Hiyoshi M, Mwimanzi P, Ueno T, Adachi A, et al. (2011) The Identification of a Small Molecule Compound That Reduces HIV-1 Nef-Mediated Viral Infectivity Enhancement. PLoS ONE 6(11): e27696. doi:10.1371/journal.pone.0027696

**Editor:** Olivier Schwartz, Institut Pasteur, France

**Received:** March 21, 2011; **Accepted:** October 22, 2011; **Published:** November 15, 2011

**Copyright:** © 2011 Chutiwitoonchai et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** This research was supported by the grant from the Global COE program "Global Education and Research Center Aiming at the control of AIDS", launched as a project commissioned by the Ministry of Education, Science, Sports, and Culture, Japan. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The authors have declared that no competing interests exist.

\* E-mail: ssuzu06@kumamoto-u.ac.jp

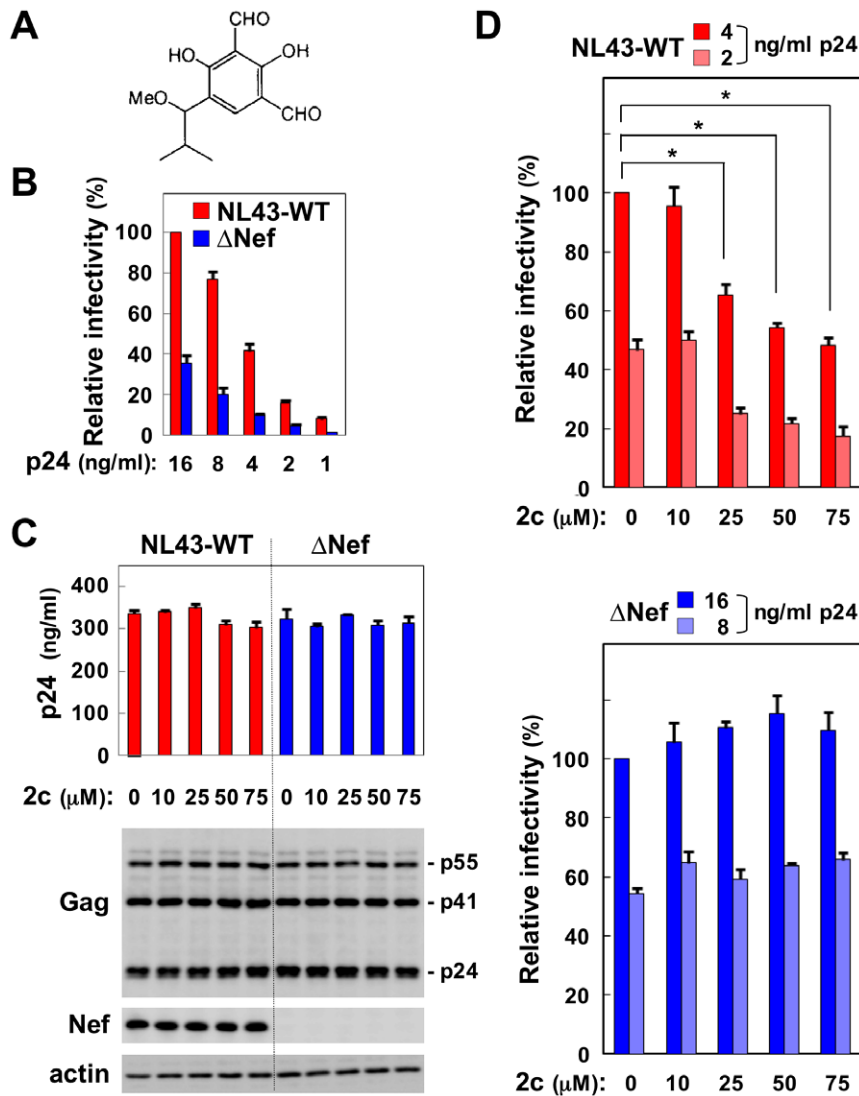
## Introduction

Nef is a 25- to 30-kDa protein with no catalytic activity encoded by the HIV-1 genome [1–4]. Studies of HIV-1-infected patients have demonstrated Nef to be a critical determinant of the progression to AIDS: HIV-1 strains without an intact *nef* gene were frequently isolated from non-progressive long-term survivors [5,6]. A subsequent study of HIV-1 transgenic mice confirmed the pathogenic activity of Nef: targeted expression of the entire coding sequence of HIV-1 in CD4<sup>+</sup> T cells and macrophages caused a severe AIDS-like disease in mice, which was completely abolished by disruption of the *nef* gene [7].

Nef is multifunctional. For instance, it accelerates the endocytosis of CD4 [8,9], the primary entry receptor for HIV-1, which allows efficient viral release from host cells [1–4]. Nef also reduces the surface expression of MHC I through multiple mechanisms [10–13], which diminishes the recognition of infected cells by CTL [1–4]. Nef is also known to activate the Src kinase Hck [14–16], which causes an impaired macrophage response to the cytokine M-CSF [17,18] or triggers cell fusion of HIV-1-infected macrophages [19]. Another hallmark function of Nef is the

enhancement of the intrinsic infectivity of progeny viruses. This function of Nef is independent of CD4 downregulation and requires the presence of Nef in viral producer cells [20–23]. Moreover, this function appears to depend on an early step of the target cell infection process, as Nef is dispensable for the infectivity of HIV-1 pseudotyped with vesicular stomatitis virus glycoprotein VSV-G [24,25]. However, Nef does not affect viral assembly or maturation, and it is still unclear how Nef enhances viral infectivity [26].

Thus far, only a few chemical compounds that interfere with the functions of Nef have been identified. Among them, a series of guanidine alkaloid analogs were found to be too toxic for cell-based assays [27]. A unique diphenylfuropyrimidine and its analogs were identified to be strong inhibitors of the Nef-dependent activation of Hck, but their primary target seemed to be Hck not Nef [28]. In contrast, the chemical compounds D1 and 2c directly target Nef. Betzi et al. identified D1 and showed that it reduced Nef-mediated MHC I, but not CD4, downregulation in a dose-dependent manner [29]. Subsequently, we identified 2c, the structure of which is distinct from that of D1, and showed that it almost completely inhibited the Nef-dependent activation of Hck

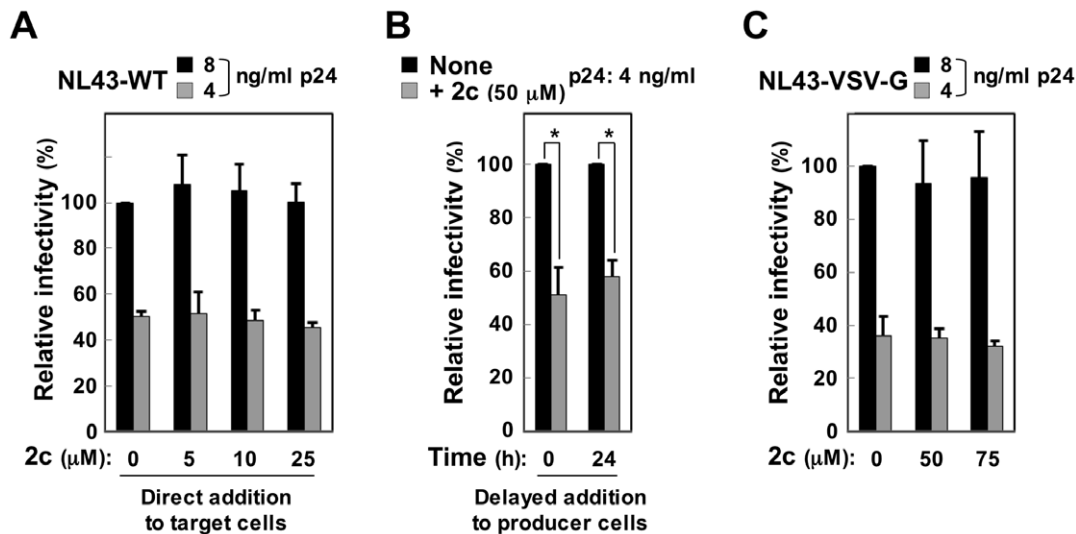


**Figure 1. The effect of 2c on the infectivity of NL43 wild-type and Nef-defective mutant viruses.** (A) The chemical structure of 2c. (B) The infectivity of the NL43 wild-type (WT) and Nef-defective mutant ( $\Delta$ Nef) viruses to the target TZM-bl cells was compared by varying the concentration of p24 Gag protein and is expressed as a percentage of the value for the sample on the far left. Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of two independent experiments with similar results. (C) 2c was added to 293 cells producing NL43-WT or  $\Delta$ Nef viruses at the indicated concentrations for 2 days, and the concentration of p24 Gag protein in the cell supernatants was determined by ELISA (bar graph). Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of two independent experiments with similar results. Alternatively, the producer cells were lysed and analyzed for the expression of Gag and Nef by Western blotting (lower blots). The actin blot was used as a loading control. (D) The infectivity of NL43-WT (upper) or  $\Delta$ Nef viruses (lower) produced by 293 cells in the absence or presence of the indicated concentrations of 2c was determined using TZM-bl cells as the target cells. The WT and  $\Delta$ Nef viruses were inoculated by changing the p24 concentration (2 or 4 ng/ml and 8 or 16 ng/ml for the WT and  $\Delta$ Nef viruses, respectively) so that the two viruses were similarly infective to the target cells. Infectivity is expressed as a percentage of the value for the sample on the far left. Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of three independent experiments with similar results. \* $p < 0.05$ . doi:10.1371/journal.pone.0027696.g001

[30] and significantly reduced Nef-mediated MHC I, but not CD4, downregulation [31]. The fact that 2c has the inhibitory effect on MHC I downregulation and Hck activation, but not on CD4 downregulation, agrees with the finding that MHC I downregulation and Hck activation are mediated by overlapping motifs or amino acids of Nef, which are distinct from those required for CD4 downregulation [3,9,14,18]. However, none of these compounds have been tested for their ability to interfere with the enhancement of viral infectivity by Nef.

In contrast to its requirement for elevated *in vivo* viral load [5,6], Nef is not essential for viral replication in *ex vivo* cell

cultures. Nonetheless, Nef significantly enhances viral replication in primary CD4<sup>+</sup> T cells and macrophages that have been exposed to HIV-1 prior to their stimulation with mitogens [32,33], a function of Nef that is likely determined by enhancement of the initial infection with cell-free HIV-1 [34]. In this regard, a compound that can reduce viral infectivity would be a valuable chemical probe for revealing the underlying mechanism of this function of Nef. In this study, we identified 2c as the first small compound that has an inhibitory effect on Nef-mediated HIV-1 infectivity enhancement and reported its inhibitory mechanism.



**Figure 2. Several features of the activity of 2c on viral infectivity.** (A) 2c was added to the target TZM-bl cells at the indicated concentrations together with the NL43 wild-type (WT) viruses produced in the absence of 2c. The amount of p24 inoculated was 4 or 8 ng/ml. The infectivity is expressed as a percentage of the value for the sample on the far left. (B) 2c (50 μM) or the control DMSO was added to the producer 293 cells immediately after transfection (0 h) or 24 h after transfection of the NL43 WT plasmid. The infectivity of the viruses was determined using TZM-bl cells and is expressed as a percentage of the value for the sample on the far left. The amount of p24 inoculated was 4 ng/ml. (C) 2c (50 or 75 μM) or the control DMSO was added to the producer 293 cells immediately after co-transfection of Env-defective NL43 plasmid and VSV-G expression plasmid. The infectivity of the pseudotyped viruses was determined using TZM-bl cells and is expressed as a percentage of the value for the sample on the far left. The amount of p24 inoculated was 4 or 8 ng/ml. (A–C) Data are shown as the mean ± SD of triplicate assays and are representative of two independent experiments with similar results. \* $p < 0.05$ . doi:10.1371/journal.pone.0027696.g002

## Results and Discussion

### 2c reduces the infectivity of wild-type HIV-1

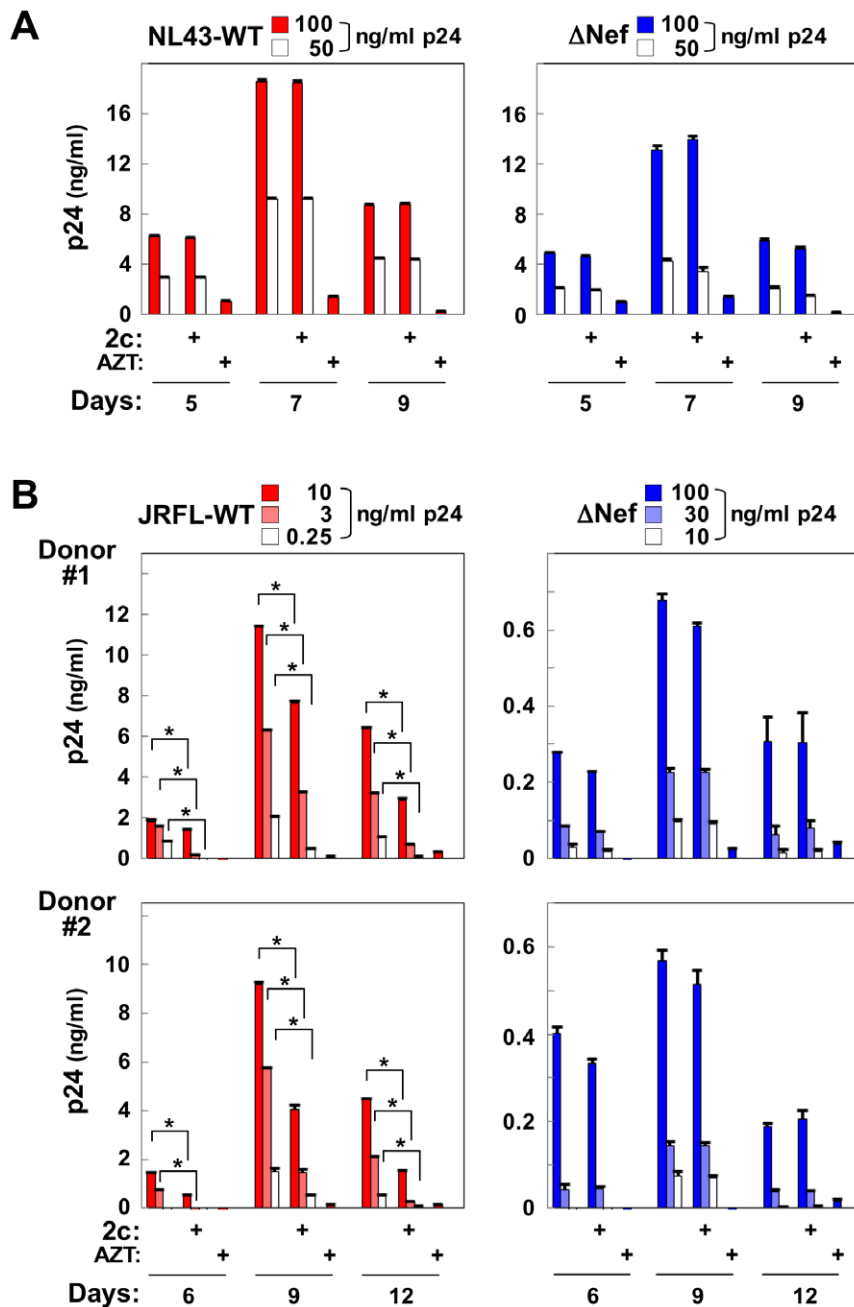
We assessed the effect of the compound 2c (Fig. 1A) on Nef-mediated infectivity enhancement using a standard single-round of replication assay [21–23]. HIV-1 viruses were prepared by transfecting HIV-1 proviral clones into 293 cells (producer cells), and infectivity was analyzed by inoculating TZM-bl cells (target cells) with defined amounts of p24 Gag protein of the resultant viruses. We first used the proviral clone NL43 and a Nef-defective mutant ( $\Delta$ Nef) and confirmed that the infectivity of the  $\Delta$ Nef viruses was lower than that of the NL43 wild-type (WT) viruses (Fig. 1B). When added to the producer 293 cells, 2c did not affect the production of WT or  $\Delta$ Nef viruses, even at a high concentration such as 75 μM (Fig. 1C): there was no significant difference in the supernatant p24 Gag protein concentration (upper graph) or the processing of the Gag polypeptide in the cells (lower blots) between the control and 2c-treated cells. However, we found that the infectivity of the WT viruses produced in the presence of 2c was significantly lower than that of the control viruses (Fig. 1D, upper). An inhibitory effect of 2c was detectable at a minimal concentration of 25 μM. Importantly, no such inhibition was observed for the  $\Delta$ Nef viruses, even at a high 2c concentration (75 μM) (Fig. 1D, lower). In the experiment shown in Fig. 1D, WT and  $\Delta$ Nef viruses were inoculated into TZM-bl cells, and the concentration of p24 was adjusted (2 or 4 ng/ml and 8 or 16 ng/ml for WT and  $\Delta$ Nef viruses, respectively) so that the two viruses were similarly infective to the target cells (see Fig. 1B). As the supernatant of proviral plasmid-transfected 293 cells was used as a viral stock, 2c was also present in the culture of target cells (<5 μM). However, 2c did not reduce the infectivity when added to the target cells at a high concentration (10 or 25 μM) together with WT viruses produced in the absence of 2c (Fig. 2A), suggesting that the presence of 2c in the producer cells was

essential for its inhibitory effect. Although 2c was added to the producer cells immediately after transfection in Fig. 1D, the inhibitory effect was also observed when 2c was added 24 h after transfection (Fig. 2B). Importantly, 2c did not show any inhibitory effect on the infectivity of Nef<sup>+</sup> HIV-1 viruses pseudotyped with VSV-G (Fig. 2C), which was consistent with the finding that Nef was dispensable for the infectivity of VSV-G-pseudotyped HIV-1 [24,25]. Therefore, these results indicated that 2c specifically reduced the infectivity of the wild-type NL43 viruses produced in the presence of Nef.

We also assessed the effect of 2c on viral replication. 2c decreased by half in the number of viable peripheral blood mononuclear cells after 9 days when used at 50 μM (data not shown). On the other hand, 2c at the same concentration showed no detectable toxicity to 293, TZM-bl, Jurkat T cells and macrophages (data not shown). We therefore used Jurkat and macrophages as target cells. As previously reported [28], the replication of HIV-1 NL43 was independent of Nef in Jurkat T cells (Fig. 3A). Accordingly, 2c failed to inhibit viral replication in the cells (Fig. 3A). However, WT JRFL viruses replicated more efficiently than  $\Delta$ Nef viruses in monocyte-derived macrophages, and 2c significantly reduced the replication of WT viruses (Fig. 3B). The result further supported the idea that the primary target of 2c was Nef.

### The inhibitory effect of 2c requires the proline-rich PxxP motif of Nef

Next, we tested the inhibitory activity of 2c on the infectivity of NL43 viruses with point mutations in Nef; i.e., R77A, K82A, D86A, F90A, or G119L [35]. As shown, 2c reduced the infectivity of all these viruses, although to a varying degree (Fig. 4A). Interestingly, the intrinsic infectivity of the NL43-G119L viruses was shown to be low [35] (also see Fig. 4A), but 2c further reduced the infectivity of the mutant viruses to the level of the  $\Delta$ Nef viruses

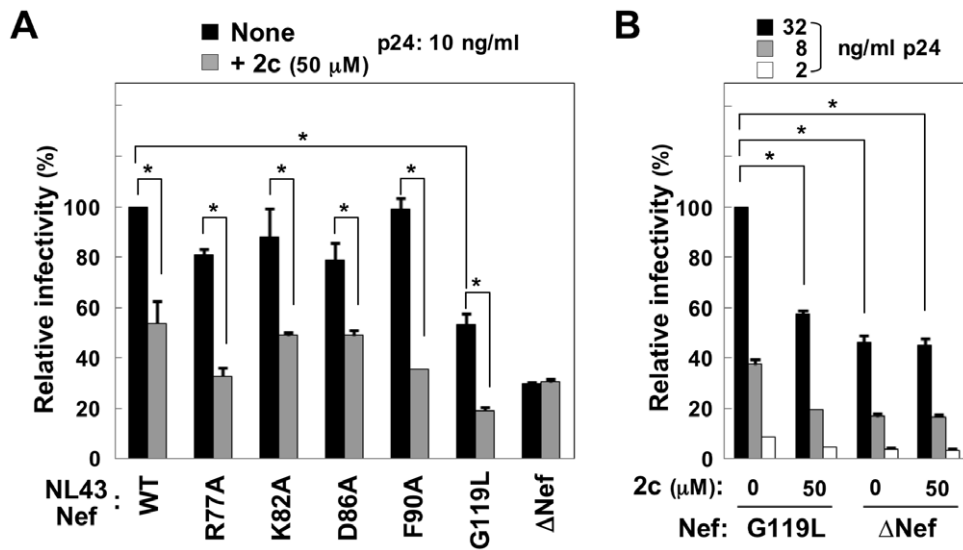


**Figure 3. The effect of 2c on the replication of HIV-1.** (A) Jurkat cells were infected with either the NL43 wild-type (WT) or Nef-defective ( $\Delta$ Nef) viruses at the indicated concentrations of p24, and cultured in the presence (50  $\mu$ M) or absence of 2c. AZT was also used at 5  $\mu$ M. The concentration of p24 in the supernatants (at day 5, 7 or 9) was determined by ELISA. Data are shown as the mean  $\pm$ SD of triplicate assays and are representative of two independent experiments with similar results. (B) Peripheral blood monocyte-derived macrophages were obtained from two different donors, infected with either the JRFL wild-type (WT) or Nef-defective ( $\Delta$ Nef) viruses at the indicated concentrations of p24, and cultured in the presence (50  $\mu$ M) or absence of 2c. AZT was also used at 5  $\mu$ M. The concentration of p24 in the supernatants (at day 6, 9 or 12) was determined by ELISA. Data are shown as the mean  $\pm$ SD of triplicate assays. \* $p$ <0.05. doi:10.1371/journal.pone.0027696.g003

(Fig. 4B). This result supported the conclusion that 2c reduced the infectivity of the NL43 viruses in a Nef-dependent manner.

The dileucine motif of Nef ( $^{164}$ LL $^{165}$ ) that is required for CD4 downregulation is also required for the enhancement of infectivity [3,36]. However, it was unlikely that the inhibitory activity of 2c was mediated through the motif, as 2c did not inhibit CD4 downregulation [31]. On the other hand, Nef has another characteristic motif; i.e., a proline-rich PxxP motif, and the

substitution of the proline residues for alanine residues (AxxA) is known to result in reduced viral infectivity [3]. Thus, we tested whether 2c further reduced the infectivity of Nef-AxxA viruses as it did with G119L mutant viruses (see Fig. 4B). To test whether 2c is also effective against Nef derived from an additional HIV-1 strain, we used an HIV-1 JRFL construct in which *nef* gene was replaced with that of the SF2 strain Nef or its AxxA mutant [30] in the subsequent experiments. First, as expected, the infectivity of the



**Figure 4. The effect of 2c on the infectivity of NL43 viruses with point amino acid mutations in Nef.** (A) The infectivity of the indicated NL43 viruses produced by 293 cells in the absence or presence of 50  $\mu$ M 2c was determined using TZM-bl cells as the target cells and is expressed as a percentage of the value for the sample on the far left. The amount of p24 inoculated was 10 ng/ml. Wild-type (WT), Nef-defective ( $\Delta$ Nef), or viruses with the indicated amino acid point mutations in Nef (R77A, K82A, D86A, F90A, or G119L) were used. (B) The infectivity of NL43 viruses with the G119L mutation in Nef or  $\Delta$ Nef viruses produced by 293 cells in the absence or presence of 50  $\mu$ M 2c was determined using TZM-bl cells as the target cells and is expressed as a percentage of the value for the sample on the far left. The amount of p24 inoculated was 2, 8, or 32 ng/ml. (A, B) Data are shown as the mean  $\pm$ SD of triplicate assays and are representative of two independent experiments with similar results. \* $p < 0.05$ . doi:10.1371/journal.pone.0027696.g004

Nef-AxxA viruses was lower than that of the wild-type (WT) viruses, although it was still higher than that of the  $\Delta$ Nef viruses (Fig. 5A). As was the case with the NL43 viruses (see Fig. 1C), 2c did not affect viral production in the JRFL-SF2 Nef viruses (Fig. 5B): there was no change in the amount of p24 Gag protein in the supernatants (upper graph), the processing of the Gag polyprotein, or the expression of Nef or another viral protein, Vif, (lower blots) between the control and 2c-treated cells. Moreover, as was the case with the NL43 viruses (see Fig. 1D), 2c significantly reduced the infectivity of the produced JRFL-SF2 Nef WT viruses, but not that of the  $\Delta$ Nef viruses (Fig. 5C). However, we found that 2c minimally affected the infectivity of the Nef-AxxA mutant viruses (Fig. 5C, middle), which was in contrast with the finding that it further reduced the infectivity of the Nef-G119L mutant viruses (see Fig. 4B). These results suggested that the inhibitory activity of 2c is mediated, at least in part, through the proline-rich motif of Nef.

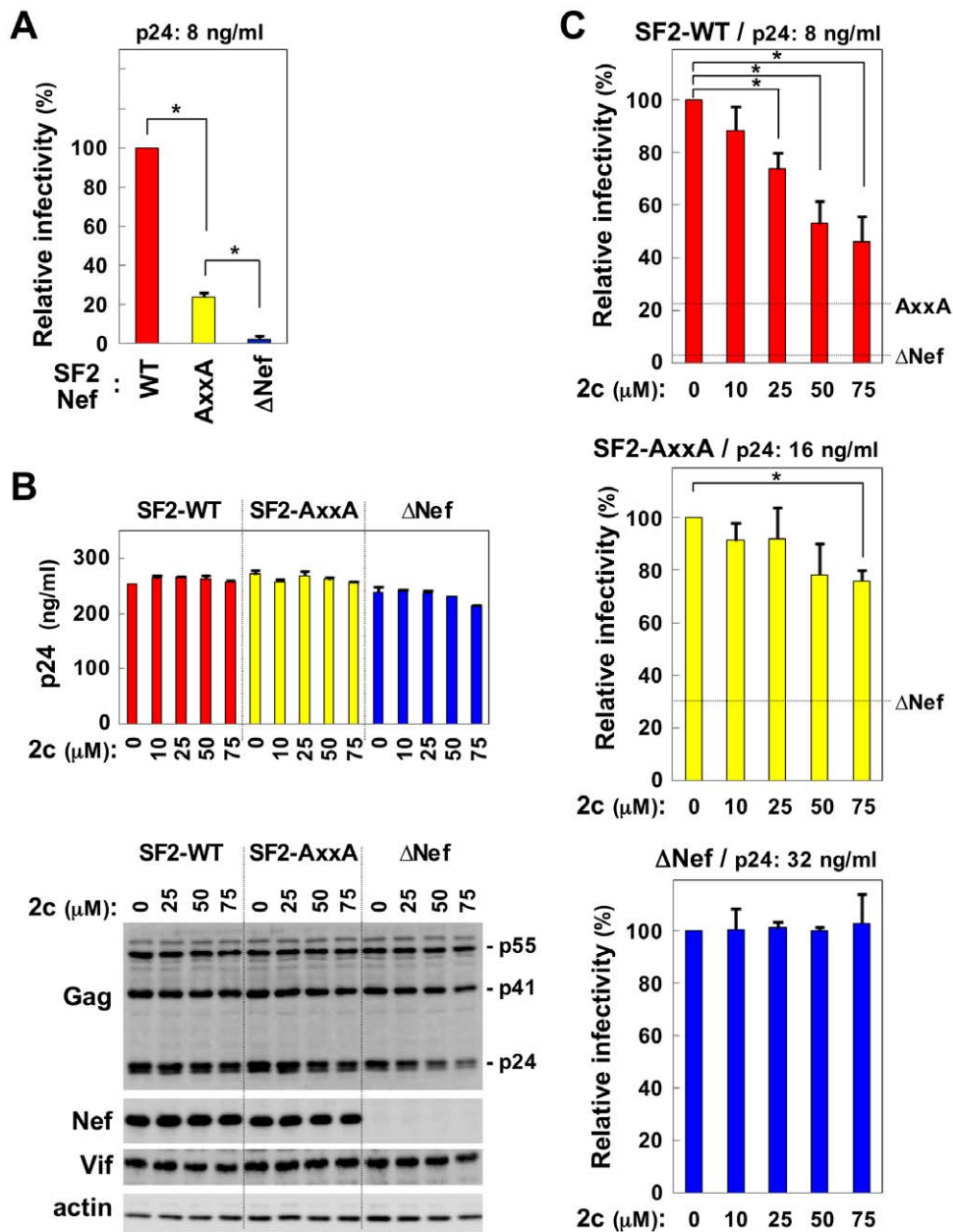
### 2c binds directly to Nef in a similar manner to the Hck SH3 domain

Although 2c was the first small molecule to be found to reduce the Nef-mediated infectivity of HIV-1, the overexpression of mutant forms of Hck in viral producer cells was also reported to result in reduced viral infectivity [37]. Hck is a cellular tyrosine kinase, and its SH3 domain has been shown to bind to Nef with high affinity [14–16], although its pathological significance is not yet understood. It is also known that the SH3 domain forms an intra-molecular interaction with the linker region of Hck [15,16] (also see Fig. 6A). Thus, the SH3 domain of mutant Hck, which lacks the linker region and the subsequent kinase domain (see Fig. 6A, HckN), is devoid of the intra-molecular interaction, and is thought to more efficiently bind to Nef and thereby reduce viral infectivity. Indeed, when co-expressed with the NL43 proviral clone, HckN and HckN-R151S, which carries a mutation in its SH2 domain, but not HckN-W93F, which carries a mutation in its

SH3 domain, significantly reduced the infectivity of viruses produced from the cells (Fig. 6B).

Based on these results, we hypothesized that 2c inhibits viral infectivity in a similar manner to mutant Hck. To this end, we examined whether 2c and Hck compete to bind to Nef using an *in vitro* pull-down assay. First, we performed a pull-down assay with various combinations of GST-Nef fusion proteins (Fig. 7A) and the Hck proteins described above. As a result, we found that the wild-type (WT) NL43 Nef bound to the wild-type (WT) Hck, HckN, and HckN-R151S, but not Hck-W93F, which had a mutation in its Nef-binding SH3 domain (Fig. 7B). In contrast, the PxxP motif-disrupted AxxA mutant did not bind to any of these Hck proteins (Fig. 7B), confirming that the pull-down system specifically detected Nef-Hck binding. As the affinity of the SF2 strain Nef for Hck was higher than that of NL43 strain Nef, which was due to the different amino acid present within the PxxP motif (Figs. 7A and B, NL43 Nef-TR mutant with a T71R substitution), we used SF2 Nef in the following experiments. Among three different competitive pull-down assays, the pre-incubation of Nef with 2c most effectively inhibited the binding of Hck to Nef (Fig. 7C, right). We therefore concluded that both the Hck SH3 domain and 2c directly bind to Nef and that their binding sites overlap.

To further confirm the above-mentioned conclusion, we used a GST fusion protein containing a 20-mer peptide derived from the PxxP motif of SF2 Nef (Fig. 8A, SF2-PxxP). As shown, the observed binding of the SF2-PxxP peptides to Hck was specific, albeit weak, in comparison with that of the full-length Nef, since it was detected with the wild-type Hck, HckN, and HckN-R151S, but not with the Nef binding-deficient HckN-W93F (Fig. 8A). Importantly, 2c inhibited the binding of Hck to the Nef-PxxP peptide, and its inhibitory effect was more marked when the Nef-PxxP peptide was pre-incubated with 2c prior to its incubation with Hck (Fig. 8B). This result suggests that 2c binds to Nef, at least in part, through the region including the PxxP motif, which is consistent with the finding that unlike the wild-type viruses, the

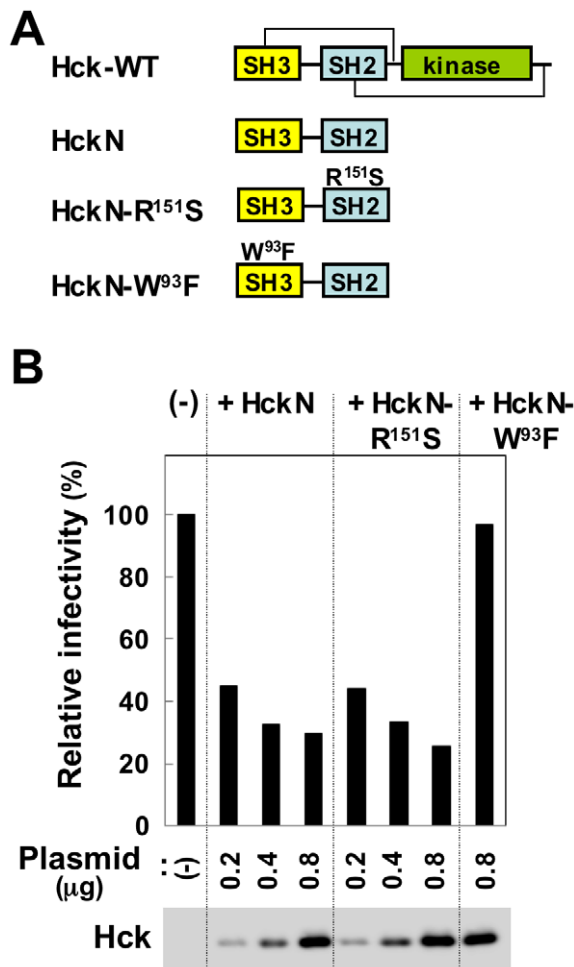


**Figure 5. The effect of 2c on the infectivity of SF2 wild-type, Nef-defective, and Nef PxxP motif-disrupted viruses.** (A) The infectivity of the SF2 wild-type (WT), Nef-defective ( $\Delta$ Nef), and Nef PxxP motif-disrupted viruses (AxxA) was compared by inoculating them into the target TZM-bl cells at a concentration of 8 ng/ml p24 and is expressed as a percentage of the value for the sample on the far left. Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of two independent experiments with similar results. \* $p < 0.05$ . (B) 2c was added to 293 cells producing SF2-WT,  $\Delta$ Nef, or AxxA viruses at the indicated concentrations for 2 days, and the concentration of p24 Gag protein in the supernatants was determined by ELISA (bar graph). Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of two independent experiments with similar results. Alternatively, the producer cells were lysed and analyzed for the expression of Gag, Nef, and Vif by Western blotting (lower blots). The actin blot was used as a loading control. (C) The infectivity of SF2-WT (top), AxxA (middle), or  $\Delta$ Nef viruses (bottom) produced by 293 cells in the absence or presence of the indicated concentrations of 2c was determined using TZM-bl cells as the target cells. The WT, AxxA, and  $\Delta$ Nef viruses were inoculated by changing the concentration of p24 (8 ng/ml, 16 ng/ml, and 32 ng/ml for WT, AxxA and  $\Delta$ Nef viruses, respectively) so that these viruses were similarly infective to the target cells. Infectivity is expressed as a percentage of the value for the sample on the far left. In the top panel, the infectivity values of the AxxA and  $\Delta$ Nef viruses produced at the same concentration of p24 (i.e., 8 ng/ml) are also shown as a reference. In the middle panel, the infectivity values of the  $\Delta$ Nef viruses produced at the same concentration of p24 (i.e., 16 ng/ml) are also shown. Data are shown as the mean  $\pm$  SD of triplicate assays and are representative of three independent experiments with similar results. \* $p < 0.05$ . doi:10.1371/journal.pone.0027696.g005

infectivity of the PxxP motif-disrupted AxxA mutant viruses was minimally affected by 2c (see Fig. 5C).

Finally, a computer-assisted simulation of the 2c-Nef docking model supported the idea that 2c binds directly to Nef and

suggested that R77, K82, A83, D86, I87, F90, Q118, and Y120 (positions are based on the sequence of NL43 strain Nef) may be responsible for this binding (Fig. 9). Among them, R77 lies within the PxxP motif ( $\text{-PVT}\underline{\text{P}}\underline{\text{Q}}\underline{\text{V}}\underline{\text{P}}\underline{\text{L}}\text{R}^{\text{77}}\underline{\text{P}}\text{-}$ , the proline residues are



**Figure 6. The effects of the overexpression of mutant forms of Hck on viral infectivity.** (A) The mutant forms of Hck used are shown schematically. HckN lacks the kinase domain and the two intramolecular interactions present in the wild-type (WT) Hck. HckN-based HckN-R151S and HckN-W93F have amino acid substitutions in their SH2 and SH3 domain, respectively. (B) The 293 cells were transfected with the NL43 wild-type proviral plasmid or co-transfected with the indicated amount of plasmid (HckN, HckN-R151S, or HckN-W93F). The infectivity of the viruses produced in the supernatants was determined using TZM-bl cells as the target cells and is expressed as a percentage of the value for the sample on the far left (bar graph). The amount of p24 inoculated was 8 ng/ml. Alternatively, the producer 293 cells were lysed and analyzed for the expression of the mutant Hck proteins by Western blotting (blot). doi:10.1371/journal.pone.0027696.g006

underlined). On the other hand, molecular modeling also identified several residues in Nef that are responsible for its binding to Hck, such as P72, P75, R77, A83, F90, W113, His116, and Y120 [38]. Among them, R77, A83, F90, and Y120 were also found in the 2c-Nef docking model (Fig. 9, underlined), supporting the finding that 2c inhibits the binding of Hck to Nef or Nef PxxP motif-derived peptides (Figs. 7C and 8B). In summary, the present study revealed that the compound 2c reduced the infectivity of HIV-1 viruses and suggested that its inhibitory activity is mediated by its direct binding to Nef.

It remains to be determined exactly how 2c reduces Nef-mediated infectivity enhancement. Given that both 2c and the Hck SH3 domain bind directly to overlapping domains of Nef and reduce viral infectivity, we speculate that their inhibitory effects

are due to the inhibition of the interaction of Nef with host proteins (Fig. 10). One of the candidates for such a host protein is p21-activated kinase 2, PAK2, the association of which depends on the Nef PxxP motif [39]. However, we did not observe any inhibitory effect of 2c on the association of Nef with PAK2 activity or its downstream effector functions (data not shown), and the Nef-PAK2 association is dispensable for the enhancement of infectivity by the viral protein [40]. Another candidate is the GTPase dynamin 2 whose interaction with Nef was implicated in enhancing viral infectivity [41]. However, again, we did not observe any significant inhibitory effect of 2c on the binding of Nef to dynamin 2, which was assessed using a co-immunoprecipitation assay (data not shown). Thus, the inhibitory activity of 2c observed in this study appears to be independent of these host proteins. The inhibitory compound 2c is a useful chemical probe for investigating the underlying molecular mechanism by which Nef enhances the infectivity of HIV-1, and in particular, for identifying the host proteins involved in the process.

Recently, a single-domain antibody (sdAb) that binds to Nef was reported [42]. Although the binding domains in Nef remained unclear, anti-Nef sdAb was also shown to reduce *in vitro* viral infectivity [42]. Therefore, to clarify whether viral infectivity enhancement by Nef accounts for the high *in vivo* viral load observed in the presence of Nef, it is necessary to test the effects of 2c, a more potent analog, and/or the anti-Nef sdAb in animal models such as HIV-1-infected humanized mice.

## Materials and Methods

### The compound 2c preparation

Some of the 2c was prepared by Kyowa Hakko Kogyo (Tokyo, Japan), as described previously [43], whilst the rest (a large quantity) was prepared by Sai Advantium Pharma (Hyderabad, India). Both preparations were dissolved in DMSO and had an equivalent inhibitory effect on HIV-1 infectivity (data not shown).

### Proviral plasmids

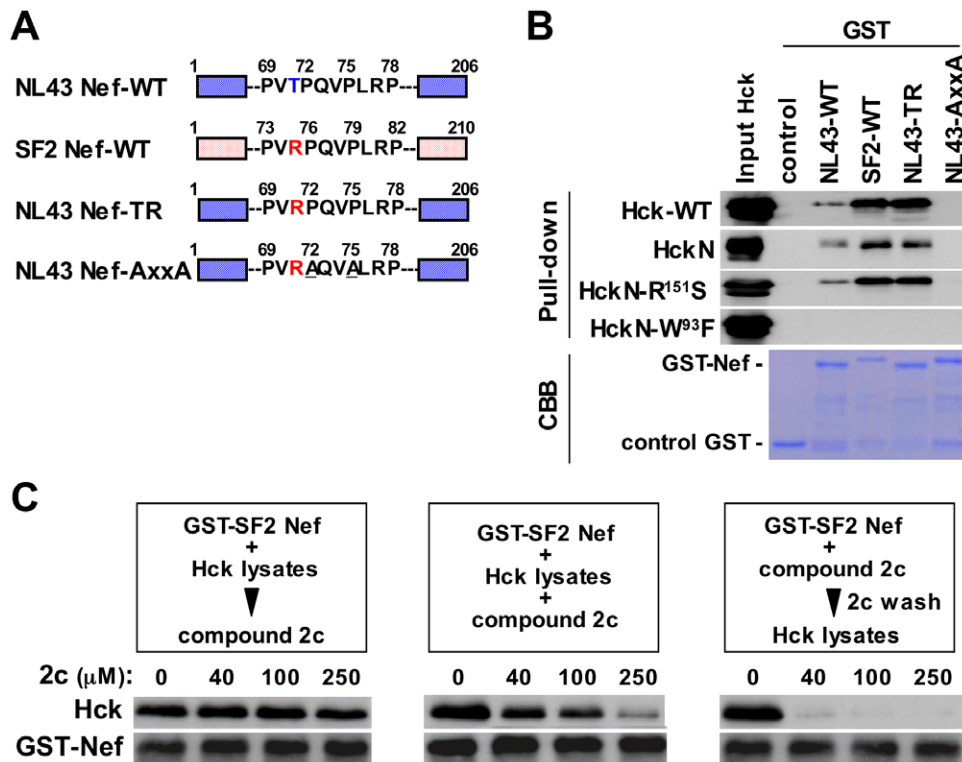
The proviral NL43 plasmid and its derivatives, which had mutations in the Nef gene ( $\Delta$ Nef, R77A, K82A, D86A, F90A, and G119L), were prepared as described previously [35]. The Env-defective mutant (pNL-Kp) and VSV-G expression plasmid were prepared as described previously [44]. The proviral JRFL plasmid was provided by Y. Koyanagi (Kyoto University, Kyoto, Japan) [45]. We also prepared the proviral JRFL plasmid, in which the Nef gene was disrupted ( $\Delta$ Nef) or replaced with the PxxP motif-disrupted AxxA mutant [30].

### Hck plasmids

The p56Hck cloned into the pcDNA3.1 vector (Invitrogen) was prepared as described previously [18]. The mutant forms of Hck cloned into the pCAGGS vector (HckN, Hck-R151S, and Hck-W93F; see Fig. 6A) were provided by M. Matsuda (Kyoto University, Kyoto, Japan) [37].

### GST fusion plasmids

The control GST and GST-Nef fusion plasmids (the wild-type NL43, NL43 Nef-TR mutant, NL43 Nef-AxxA, and the wild-type SF2; see Fig. 7) were prepared as described previously [30]. We also prepared a GST-SF2 Nef-PxxP plasmid, which expressed a 20-mer peptide derived from the PxxP motif of SF2 Nef (see Fig. 8). The cDNA containing the motif was amplified by PCR using the following primers (5'-GGATCCGTGGGTTTTCCAGT-3' and 5'-GTCGACCTATAAAGCTGCCT-3'), cloned into the pCR2.1 vector (Invitrogen), sequenced using the BigDye Terminator v3.1



**Figure 7. The effect of 2c on binding between Nef and Hck.** (A) The Nef proteins fused to GST are shown schematically. In addition to the wild-type (WT) SF2 and NL43 strain Nef, the NL43-TR mutant, which contained a T71R amino acid substitution, and another NL43 AxxA mutant, in which the PxxP motif was disrupted (P72A and P75A substitutions), were used. (B) The resins to which the control GST or indicated GST-Nef fusion proteins were bound were incubated with the lysates of 293 cells expressing the indicated Hck protein. The amount of Hck bound to the resins was determined by Western blotting (pull-down assay). To confirm the equal expression of these Hck proteins in the 293 cells, equal amounts of each cell lysate were analyzed (Input Hck). Moreover, the amounts of the GST and GST-Nef fusion proteins bound to the resins were verified by the elution from the resins followed by SDS-PAGE/Coomassie brilliant blue (CBB) staining. (C) Three different competitive pull-down assays were performed. In the experiment shown in the left panel, the resins to which the GST-SF2 Nef fusion proteins were bound were incubated with the lysates obtained from the 293 cells expressing the wild-type Hck for 3 h, and then 2c was added to the mixture at the indicated concentration. In the experiment shown in the middle panel, the resins to which the GST-SF2 Nef fusion proteins were bound were incubated with the lysates of 293 cells expressing the wild-type Hck and the indicated concentration of 2c. In the experiment shown in the right panel, the resins to which the GST-SF2 Nef fusion proteins were bound were first incubated with the indicated concentration of 2c for 4 h and then washed to remove unbound 2c. Then, the resins were incubated with the lysates of 293 cells expressing the wild-type Hck. The amount of Hck bound to the resins was determined by Western blotting (upper blots). The GST-Nef blot was used as a loading control (lower blots). Data shown are representative of two independent experiments with similar results.

doi:10.1371/journal.pone.0027696.g007

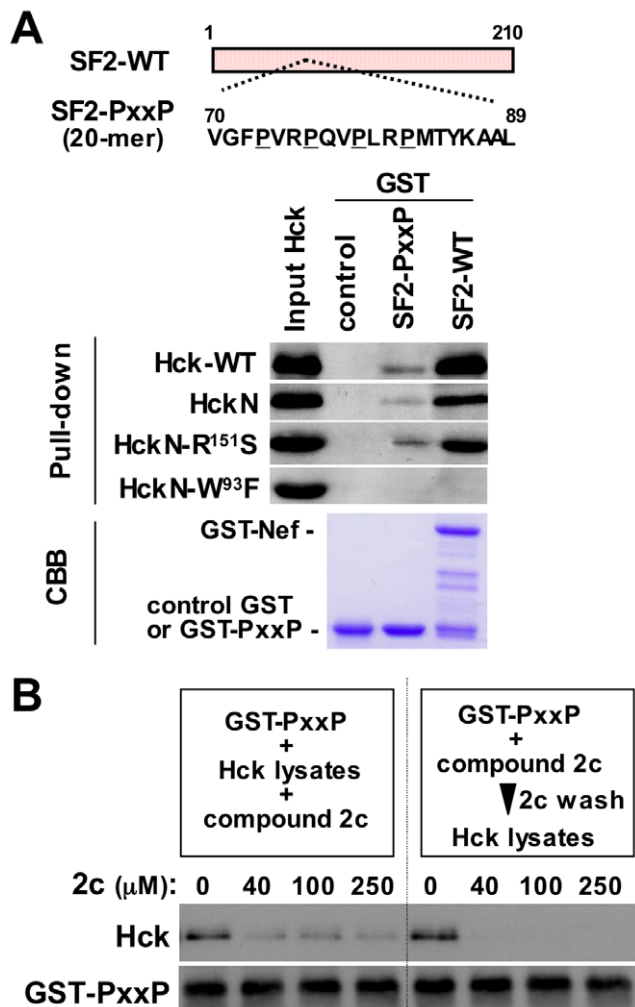
Cycle Sequencing kit (Applied Biosystems) and the ABI PRISM 3100 Genetic Analyzer (Applied Biosystems), and cloned into the pGEX-6P-1 bacterial expression vector (GE Healthcare).

### Virus preparation

HEK293 cells (Invitrogen) were maintained in DME medium supplemented with 10% FCS and used as viral producer cells. The 293 cells were seeded onto 12-well tissue culture plates at a density of  $1.8 \times 10^5$  cells/well and transfected with 1.6 μg/well of various proviral HIV-1 plasmids using 4 μl/well Lipofectamine 2000 reagent (Invitrogen). To prepare VSV-G-pseudotyped viruses (see Fig. 2C), cells were transfected with 0.5 μg/well Env-defective mutant plasmid (pNL-Kp) and 1.0 μg/well VSV-G expression plasmid. In a selected experiment (see Fig. 6B), the cells were co-transfected with 0.8 μg/well pNL43 plasmid and 0.2, 0.4, or 0.8 μg/well of one of the mutant forms of Hck (HckN, HckN-R<sup>151</sup>S, or HckN-W<sup>93</sup>F). The total amount (1.6 μg/well) of the plasmid was normalized using the pCAGGS empty vector. After 6 h of transfection, the culture medium was replaced with fresh medium, and the cells were cultured for an additional 48 h in the

presence or absence of 2c at the indicated concentrations. In a selected experiment (see Fig. 2B), 2c was added to the culture 24 h after transfection. Then, the supernatants containing the viruses were clarified by brief centrifugation, and viral production was assessed by measuring the concentration of p24 Gag protein in the supernatants using the RETROtek p24 Antigen ELISA kit (ZeptoMetrix). Viral production was also assessed by analyzing the expression of viral proteins in the cells by Western blotting. The preparation of the total cell lysates and Western blotting were performed essentially as described previously [17,18,30]. Briefly, the cells were lysed on ice with Nonidet P-40 lysis buffer (1% Nonidet P-40, 50 mM Tris, and 150 mM NaCl) containing protease inhibitors (1 mM EDTA, 1 μM PMSF, 1 μg/ml aprotinin, 1 μg/ml leupeptin, and 1 μg/ml pepstatin). Total cell lysates were then subjected to Western blotting. The antibodies used were as follows: anti-Gag (#65-004; BioAcademia, Osaka, Japan), anti-Nef (#2949; NIH AIDS Research & Reference Program), anti-Vif (#319; NIH AIDS Research & Reference Program), and anti-actin (#C-2; Santa Cruz). The detection was performed with HRP-labeled secondary antibodies (GE Health-





**Figure 8. The effect of 2c on the binding between Nef PxxP motif-containing peptides and Hck.** (A) The Nef peptide fused to GST is shown schematically. The 20 amino acid peptide derived from the PxxP motif of SF2 Nef was used (the proline residues are underlined). The resins to which the control GST, GST-SF2 Nef-PxxP peptides (SF2-PxxP), or GST-intact SF2 Nef (SF2-WT) fusion proteins were bound were incubated with the lysates of 293 cells expressing the indicated Hck protein. The amount of Hck bound to each resin was determined by Western blotting (Pull-down). To verify the equal expression of these Hck proteins in the 293 cells, equal amounts of each cell lysate were analyzed (Input Hck). Moreover, the amounts of the GST and GST-Nef fusion proteins bound to the resins were verified by eluting from the resins followed by SDS-PAGE/Coomassie brilliant blue (CBB) staining. (B) Two different competitive pull-down assays were performed. In the experiment shown in the left panel, the resins to which the GST-SF2 Nef-PxxP peptides were bound were incubated with the lysates of 293 cells expressing the wild-type Hck and the indicated concentration of 2c. In the experiment shown in the right panel, the resins to which the GST-SF2 Nef-PxxP peptides were bound were incubated with the indicated concentrations of 2c for 4 h and then washed to remove unbound 2c. Then, the resins were incubated with the lysates of 293 cells expressing the wild-type Hck. The amount of Hck bound to the resins was determined by Western blotting (upper blots). The GST-Nef blot was used as a loading control (lower blots). Data shown are representative of two independent experiments with similar results.  
doi:10.1371/journal.pone.0027696.g008

care), the Immunostar LD Western blotting detection reagent (Wako, Osaka, Japan), and an image analyzer (ImageQuant LAS 4000; GE Healthcare).

## Infectivity assay

TZM-bl cells (NIH AIDS Research & Reference Program) were maintained in DME medium supplemented with 10% FCS and used as viral target cells. TZM-bl cells were seeded onto 96-well tissue culture plates at a density of  $6 \times 10^3$  cells/well and challenged with serially diluted viruses normalized for the concentration of p24 Gag protein. The supernatant of the proviral plasmid-transfected 293 cells was used as a viral stock and diluted with DME medium containing 10% FCS and 20  $\mu\text{g}/\text{ml}$  DEAE-dextran (MP Biomedicals, Solon, OH). The diluted viruses were then added to the target cells (150  $\mu\text{l}/\text{well}$ ) overnight, and the culture medium was replaced with fresh DME medium containing 10% FCS and incubated for 48 h. In a selected experiment (see Fig. 2A), 2c was added to TZM-bl cells together with the diluted viruses. Viral infectivity was assessed by measuring the HIV-1 Tat-mediated induction of  $\beta$ -galactosidase activity in the target cells using a  $\beta$ -Galactosidase Enzyme Assay System (Promega). The absorbance of the wells was measured at 420 nm using a Multiskan microplate reader (Thermo Electron).

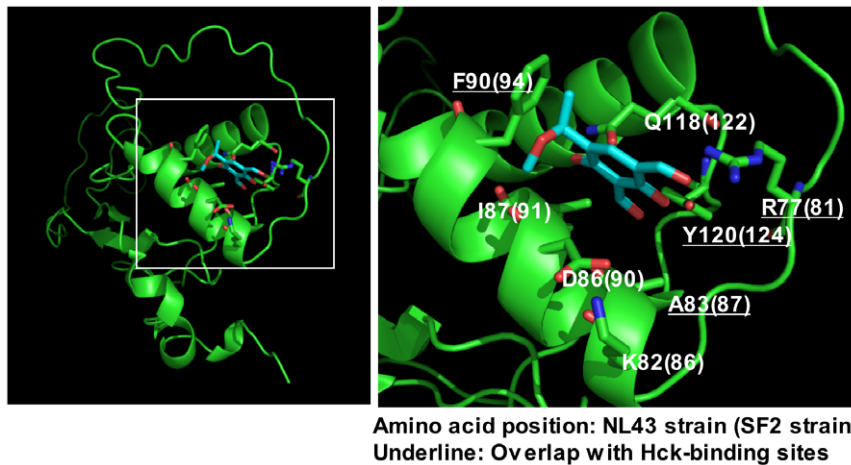
## Replication assay

The replication assay with macrophages was performed essentially as described previously [46]. Heparinized venous blood was collected from healthy donors, after informed consent was obtained in accordance with the Declaration of Helsinki. The approval for this study was obtained from the Kumamoto University Medical Ethical Committee. Mononuclear cells obtained using LSM reagent (MP Biomedicals) were suspended into RPMI1640 medium-1% FCS at  $1 \times 10^6$  cells/ml and seeded into 24-well plates. Monocytes were enriched by adherence to plates for 1 h at  $37^\circ\text{C}$ , and non-adherent cells were removed by extensive washing with PBS. Then, the adherent monocytes were differentiated into macrophages by culturing with RPMI1640-10% FCS containing 100 ng/ml rhM-CSF (a gift from Morinaga Milk Industry, Kanagawa, Japan). After 3 days, the cultures were replaced with fresh complete media and incubated for another 3 days. The purity of the day 6-macrophages prepared by this method was routinely more than 95% when assessed by the expression of CD14 (data not shown). Then, macrophages were incubated with 250  $\mu\text{l}$  of the 293 cell supernatants containing JRFL HIV-1 viruses for 2 h at  $37^\circ\text{C}$ . Either 2c or DMSO was added to the incubation together with the diluted viruses. AZT (NIH AIDS Research & Reference Program) was used as a positive control. The cells were washed twice with PBS to remove unbound viruses and cultured with RPMI1640-10% FCS containing rhM-CSF in the presence or absence of 2c or AZT. One-half of the culture media was replaced with the complete media every 3 days. The culture supernatants collected at day 6, 9 and 12 were analyzed for the concentration of p24 Gag proteins by ELISA to monitor viral replication.

Jurkat cells were also used in this study. The cell pellet ( $1 \times 10^6$  cells) were incubated with 500  $\mu\text{l}$  of the 293 cell supernatants containing NL43 viruses for 2 h at  $37^\circ\text{C}$ . Either 2c or DMSO was added to the incubation together with the diluted viruses. AZT was used as a positive control. The cells were washed twice with PBS, resuspended into 1 ml of RPMI1640-10% FCS, and cultured for 3 days in the presence or absence of 2c or AZT. Then, the culture were diluted (1/5) with RPMI1640-10% FCS, and cultured for another 2 days in the presence or absence of 2c or AZT. The concentration of p24 in the culture supernatants of day 5, 7 and 9 was analyzed as above.

## GST pull-down assay

The control GST and GST-Nef fusion proteins cloned in the pGEX-6P-1 vector were expressed in *E. coli* BL21 cells



**Figure 9. The 2c-Nef docking model.** The amino acids that are predicted to be involved in the interaction between Nef and 2c are indicated. The positions of these amino acids in the NL43 strain and SF2 strain are shown. The amino acids predicted to interact with the Hck SH3 domain are underlined [38].

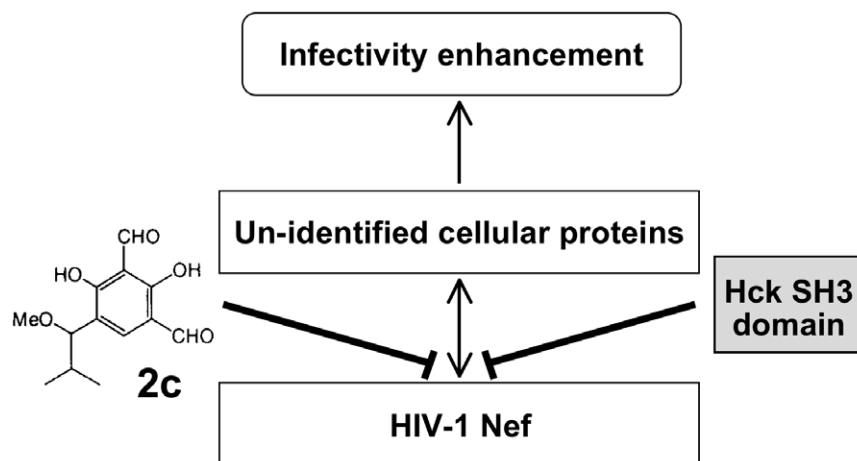
doi:10.1371/journal.pone.0027696.g009

(GE Healthcare). The cells were grown in LB medium containing 50 µg/ml ampicillin, before being induced with 1 µM IPTG (Sigma). The expression-induced cells were harvested and lysed with BugBuster Protein Extraction Reagent containing 1 U/ml rLysozyme and 25 U/ml Benzonase Nuclease (all from Novagen). The cleared lysates were then incubated with GST-Bind Resin (Novagen). After extensive washing with GST Bind/Wash Buffer (Novagen), the resin was incubated with the total cell lysate of the 293 cells transfected with the expression plasmid for Hck for 12 h. In the competitive pull-down assay, we employed the following 3 protocols: (1) the concurrent addition of 2c and Hck-containing lysates to the GST-Nef-bound resin, (2) the addition of Hck-containing lysates for 3 h followed by the addition of 2c, (3) the addition of 2c at the indicated concentrations for 4 h followed by the addition of the Hck-containing lysates. The incubation of the above mixtures was carried out at 4°C in Nonidet P-40 lysis buffer (1% Nonidet P-40, 50 mM Tris, and 150 mM NaCl) containing protease inhibitors (1 mM EDTA, 1 µM PMSF, 1 µg/ml aprotinin, 1 µg/ml leupeptin, and 1 µg/ml pepstatin). After extensive re-washing with complete Nonidet P-40 lysis buffer,

the resin was boiled with SDS-PAGE sample buffer, and the eluates were analyzed for the presence of Hck by Western blotting with anti-Hck antibodies (clone 18; Transduction Laboratories).

#### The 2c-Nef docking model

We predicted the complex structures of Nef and 2c by homology modeling and docking simulation using the Molecular Operating Environment (MOE) ver. 2007.09. (Chemical Computing Group, Canada). First, homology modeling [47–49] was used to construct the model structure of HIV-1 Nef SF2 strain using its NMR structure (PDB code: 2NEF) [50] as a template. During the modeling, energy calculations were performed with the AMBER ff99 force field [51] and the GB/VI implicit solvent energy function [52]. Next, docking simulation of 2c with the homology model of Nef was achieved with the ASEDock module [53]. The initial structure of 2c was generated with the Molecular Builder module. Then, we searched for the binding site of 2c with the Site-Finder module. During the simulation, the energy calculations were performed with the MMFF94x force field [54,55] and the GB/VI implicit solvent energy function [52].



**Figure 10. A model of the inhibitory effect of 2c.** Both 2c and the Hck SH3 domain bind directly to Nef and reduce viral infectivity, probably by inhibiting the interaction of Nef with an unidentified cellular protein(s).

doi:10.1371/journal.pone.0027696.g010

During the docking simulation, movement of the main chain atoms around 4.5 Å of the ligand binding site in Nef was restrained with a harmonic potential of 100 kcal/mol/Å<sup>2</sup>, while the atoms in compound 2c were not constrained. In this study, the structure with the lowest score was selected for the model.

### Statistical analysis

The statistical significance of differences between assay groups was determined using Mann-Whitney *U* test. *p* values less than 0.05 were considered significant.

### References

- Fackler OT, Baur AS (2002) Live and let die: Nef functions beyond HIV replication. *Immunity* 16: 493–497.
- Peterlin BM, Trono D (2003) Hide, shield and strike back: how HIV-infected cells avoid immune eradication. *Nat Rev Immunol* 3: 97–107.
- Foster JL, Garcia JV (2008) HIV-1 Nef: at the crossroads. *Retrovirology* 5: 84.
- Malim MH, Emerman M (2008) HIV-1 accessory proteins—ensuring viral survival in a hostile environment. *Cell Host Microbe* 3: 388–398.
- Deacon NJ, Tsykin A, Solomon A, Smith K, Ludford-Menting M, et al. (1995) Genomic structure of an attenuated quasi species of HIV-1 from a blood transfusion donor and recipients. *Science* 270: 988–991.
- Kirchhoff F, Greenough TC, Bretler DB, Sullivan JL, Desrosiers RC (1995) Brief report: absence of intact nef sequences in a long-term survivor with nonprogressive HIV-1 infection. *N Engl J Med* 332: 228–232.
- Hanna Z, Kay DG, Rebai N, Guimond A, Jothy S, et al. (1998) Nef harbors a major determinant of pathogenicity for an AIDS-like disease induced by HIV-1 in transgenic mice. *Cell* 95: 163–175.
- Garcia JV, Miller AD (1991) Serine phosphorylation-independent downregulation of cell-surface CD4 by nef. *Nature* 350: 508–511.
- Schaefer MR, Wonderlich ER, Roeth JF, Leonard JA, Collins KL (2008) HIV-1 Nef targets MHC-I and CD4 for degradation via a final common β-COP-dependent pathway in T cells. *PLoS Pathog* 4: e1000131.
- Schwartz O, Maréchal V, Le Gall S, Lemonnier F, Heard JM (1996) Endocytosis of major histocompatibility complex class I molecules is induced by the HIV-1 Nef protein. *Nat Med* 2: 338–342.
- Greenberg ME, Iafrate AJ, Skowronski J (1998) The SH3 domain-binding surface and an acidic motif in HIV-1 Nef regulate trafficking of class I MHC complexes. *EMBO J* 17: 2777–2789.
- Williams M, Roeth JF, Kasper MR, Filzen TM, Collins KL (2005) Human immunodeficiency virus type 1 Nef domains required for disruption of major histocompatibility complex class I trafficking are also necessary for coprecipitation of Nef with HLA-A2. *J Virol* 79: 632–637.
- Singh RK, Lau D, Noviello CM, Ghosh P, Guatelli JC (2009) An MHC-I cytoplasmic domain/HIV-1 Nef fusion protein binds directly to the μ subunit of the AP-1 endosomal coat complex. *PLoS One* 4: e8364.
- Saksela K, Cheng G, Baltimore D (1995) Proline-rich (PxxP) motifs in HIV-1 Nef bind to SH3 domains of a subset of Src kinases and are required for the enhanced growth of Nef<sup>+</sup> viruses but not for downregulation of CD4. *EMBO J* 14: 484–491.
- Moarefi I, LaFevre-Bernt M, Sicheri F, Huse M, Lec CH, et al. (1997) Activation of the Src-family tyrosine kinase Hck by SH3 domain displacement. *Nature* 385: 650–653.
- Lerner EC, Smithgall TE (2002) SH3-dependent stimulation of Src-family kinase autophosphorylation without tail release from the SH2 domain in vivo. *Nat Struct Biol* 9: 365–369.
- Suzu S, Harada H, Matsumoto T, Okada S (2005) HIV-1 Nef interferes with M-CSF receptor signaling through Hck activation and inhibits M-CSF bioactivities. *Blood* 105: 3230–3237.
- Hiyoshi M, Suzu S, Yoshidomi Y, Hassan R, Harada H, et al. (2008) Interaction between Hck and HIV-1 Nef negatively regulates cell surface expression of M-CSF receptor. *Blood* 111: 243–250.
- Vérollet C, Zhang YM, Le Cabec V, Mazzolini J, Charrière G, et al. (2010) HIV-1 Nef triggers macrophage fusion in a p61Hck- and protease-dependent manner. *J Immunol* 184: 7030–7039.
- Chowers MY, Spina CA, Kwoh TJ, Fitch NJ, Richman DD, et al. (1994) Optimal infectivity in vitro of human immunodeficiency virus type 1 requires an intact nef gene. *J Virol* 68: 2906–2914.
- Chowers MY, Pandori MW, Spina CA, Richman DD, Guatelli JC (1995) The growth advantage conferred by HIV-1 nef is determined at the level of viral DNA formation and is independent of CD4 downregulation. *Virology* 212: 451–457.
- Aiken C, Trono D (1995) Nef stimulates human immunodeficiency virus type 1 proviral DNA synthesis. *J Virol* 69: 5048–5056.
- Goldsmith MA, Warmerdam MT, Atchison RE, Miller MD, Greene WC (1995) Dissociation of the CD4 downregulation and viral infectivity enhancement functions of human immunodeficiency virus type 1 Nef. *J Virol* 69: 4112–4121.
- Aiken C (1997) Pseudotyping human immunodeficiency virus type 1 (HIV-1) by the glycoprotein of vesicular stomatitis virus targets HIV-1 entry to an endocytic pathway and suppresses both the requirement for Nef and the sensitivity to cyclosporin A. *J Virol* 71: 5871–5877.
- Chazal N, Singer G, Aiken C, Hammarskjöld ML, Rekosh D (2001) Human immunodeficiency virus type 1 particles pseudotyped with envelope proteins that fuse at low pH no longer require Nef for optimal infectivity. *J Virol* 75: 4014–4018.
- Laguette N, Benichou S, Basmaciogullari S (2009) Human immunodeficiency virus type 1 Nef incorporation into virions does not increase infectivity. *J Virol* 83: 1093–1104.
- Olaszewski A, Sato K, Aron ZD, Cohen F, Harris A, et al. (2004) Guanidine alkaloid analogs as inhibitors of HIV-1 Nef interactions with p53, actin, and p56lck. *Proc Natl Acad Sci U S A* 101: 14079–14084.
- Emert-Sedlak L, Kodama T, Lerner EC, Dai W, Foster C, et al. (2009) Chemical library screens targeting an HIV-1 accessory factor/host cell kinase complex identify novel antiretroviral compounds. *ACS Chem Biol* 4: 939–947.
- Betzi S, Restouin A, Opi S, Arold ST, Parrot I, et al. (2007) Protein-protein interaction inhibition (2P2I) combining high throughput and virtual screening: Application to the HIV-1 Nef protein. *Proc Natl Acad Sci U S A* 104: 19256–19261.
- Hassan R, Suzu S, Hiyoshi M, Takahashi-Makise N, Ueno T, et al. (2009) Dysregulated activation of a Src tyrosine kinase Hck at the Golgi disturbs N-glycosylation of a cytokine receptor Fms. *J Cell Physiol* 221: 458–468.
- Dikeakos JD, Atkins KM, Thomas L, Emert-Sedlak L, Byeon JJ, et al. (2010) Small molecule inhibition of HIV-1-induced MHC-I downregulation identifies a temporally regulated switch in Nef action. *Mol Biol Cell* 21: 3279–3292.
- Miller MD, Warmerdam MT, Gaston I, Greene WC, Feinberg MB (1994) The human immunodeficiency virus-1 nef gene product: a positive factor for viral infection and replication in primary lymphocytes and macrophages. *J Exp Med* 179: 101–113.
- Spina CA, Kwoh TJ, Chowers MY, Guatelli JC, Richman DD (1994) The importance of nef in the induction of human immunodeficiency virus type 1 replication from primary quiescent CD4 lymphocytes. *J Exp Med* 179: 115–123.
- Haller C, Tibroni N, Rudolph JM, Grosse R, Fackler OT (2011) Nef does not inhibit F-actin remodelling and HIV-1 cell-cell transmission at the T lymphocyte virological synapse. *Eur J Cell Biol* in press.
- Jere A, Fujita M, Adachi A, Nomaguchi M (2010) Role of HIV-1 Nef protein for virus replication in vitro. *Microbes Infect* 12: 65–70.
- Madrid R, Janvier K, Hitchin D, Day J, Coleman S, et al. (2005) Nef-induced alteration of the early/recycling endosomal compartment correlates with enhancement of HIV-1 infectivity. *J Biol Chem* 280: 5032–5044.
- Tokunaga K, Kiyokawa E, Nakaya M, Otsuka N, Kojima A, et al. (1998) Inhibition of human immunodeficiency virus type 1 virion entry by dominant-negative Hck. *J Virol* 72: 6257–6259.
- Choi HJ, Smithgall TE (2004) Conserved residues in the HIV-1 Nef hydrophobic pocket are essential for recruitment and activation of the Hck tyrosine kinase. *J Mol Biol* 343: 1255–68.
- Rauch S, Pulkkinen K, Saksela K, Fackler OT (2008) Human immunodeficiency virus type 1 Nef recruits the guanine exchange factor Vav1 via an unexpected interface into plasma membrane microdomains for association with p21-activated kinase 2 activity. *J Virol* 82: 2918–2929.
- Schindler M, Rajan D, Specht A, Ritter C, Pulkkinen K, Saksela K, et al. (2007) Association of Nef with p21-activated kinase 2 is dispensable for efficient human immunodeficiency virus type 1 replication and cytopathicity in ex vivo-infected human lymphoid tissue. *J Virol* 81: 13005–13014.
- Pizzato M, Helander A, Popova E, Calistri A, Zamborlini A, et al. (2007) Dynamin 2 is required for the enhancement of HIV-1 infectivity by Nef. *Proc Natl Acad Sci U S A* 104: 6812–6817.
- Bouchet J, Basmaciogullari SE, Chrobak P, Stolp B, Bouchard N, et al. (2011) Inhibition of the Nef regulatory protein of HIV-1 by a single-domain antibody. *Blood* 117: 3559–3568.
- Oneyama C, Agatsuma T, Kanda Y, Nakano H, Sharma SV, et al. (2003) Synthetic inhibitors of proline-rich ligand-mediated protein-protein interaction: potent analogs of UCS15A. *Chem Biol* 10: 443–451.
- Akari H, Uchiyama T, Fukumori T, Iida S, Koyama AH, et al. (1999) Pseudotyping human immunodeficiency virus type 1 by vesicular stomatitis virus G protein does not reduce the cell-dependent requirement of vif for optimal

### Acknowledgments

We thank F. Koutaki for her secretarial assistance.

### Author Contributions

Conceived and designed the experiments: NC SS. Performed the experiments: NC MH HO HS OF PM. Analyzed the data: NC SO SS. Contributed reagents/materials/analysis tools: TU AA. Wrote the paper: NC SS.

- infectivity: functional difference between Vif and Nef. *J Gen Virol* 80: 2945–2959.
45. Koyanagi Y, Miles S, Mitsuyasu RT, Merrill JE, Vinters HV, et al. (1987) Dual infection of the central nervous system by AIDS viruses with distinct cellular tropisms. *Science* 236: 819–822.
  46. Chihara T, Suzu S, Hassan R, Chutiwitoonchai N, Hiyoshi M, et al. (2010) IL-34 and M-CSF share the receptor Fms but are not identical in biological activity and signal activation. *Cell Death Differ* 17: 1917–1927.
  47. Marti-Renom MA, Stuart AC, Fiser A, Sánchez R, Melo F, et al. (2000) Comparative protein structure modeling of genes and genomes. *Annu Rev Biophys Biomol Struct* 29: 291–325.
  48. Baker D, Sali A (2001) Protein structure prediction and structural genomics. *Science* 294: 93–96.
  49. Shirakawa K, Takaori-Kondo A, Yokoyama M, Izumi T, Matsui M, et al. (2008) Phosphorylation of APOBEC3G by protein kinase A regulates its interaction with HIV-1 Vif. *Nat Struct Mol Biol* 15: 1184–1191.
  50. Grzesiek S, Bax A, Hu JS, Kaufman J, Palmer I, et al. (1997) Refined solution structure and backbone dynamics of HIV-1 Nef. *Protein Sci* 6: 1248–63.
  51. Wang J, Cieplak P, Kollman PA (2000) How well does a restrained electrostatic potential (RESP) model perform in calculating conformational energies of organic and biological molecules? *J Comput Chem* 21: 1049–1074.
  52. Labute P (2008) The generalized Born/volume integral implicit solvent model: estimation of the free energy of hydration using London dispersion instead of atomic surface area. *J Comput Chem* 29: 1693–1698.
  53. Goto J, Kataoka R, Muta H, Hirayama N (2008) ASEDock-docking based on alpha spheres and excluded volumes. *J Chem Inf Model* 48: 583–90.
  54. Halgren TA (1999) MMFF VII. Characterization of MMFF94, MMFF94s, and other widely available force fields for conformational energies and for intermolecular-interaction energies and geometries. *J Comput Chem* 20: 720–29.
  55. Halgren TA (1999) MMFF VI. MMFF94s option for energy minimization studies. *J Comput Chem* 20: 730–48.